Evaluation of the Antioxidant Potential and Antioxidant Enzymes of Some Yemeni Grape Cultivars

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ABSTRACT

Objective: The evaluation of antioxidant activity of grape from different cultivars was conducted by several methods. Materials and Methods: Scavenging of 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical, scavenging of 2,2’-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) and formation of phosphate/molybdenum complex. Results: The phenolic content of peel and flesh from different cultivars of grapes showed concentration dependent scavenging of DPPH and ABTS radicals and formation of phosphate/molybdenum complex. The results appeared high values of correlation coefficient (R²) between phenolic content of grapes and antioxidant activity indicating the strong correlation. The antioxidant assay IC50 value was variable among different grape cultivars. The activities of antioxidant enzymes were screened in the crude extracts of the peel and flesh of the grape cultivars tested, which had high, moderate and low levels of polyphenol oxidase, peroxidase and catalase, respectively. Conclusion: The results indicated that the imported grapes retained the most antioxidant activities. Finally, this study will be supply information on the antioxidant activity of grapes that benefits grapes producers, nutritionists and consumers.

Key words: Antioxidants, Yemeni grape, Flavonoids, Antioxidant enzymes.

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INTRODUCTION

The global production of grape juices is estimated to be around 11–12 million hectoliters and the main producer and consumer countries are the United States of America, Brazil and Spain.1 in Yemen there are about 27 grape varieties reported. The major varieties include Aasmi (Red), Raziqi, Aswad (Black). Grape is an important fruit crop in Yemen. Grape consumption has been increasing since the 1970s as a consequence of rapid urban population growth and a sharp increase in demand. In a 1998 Food and Agriculture Organization (FAO) report, Yemen ranked sixth among Arab countries in grape production.2 In the past few years there has been increasing interest in the determination of suitable dietary sources of antioxidant phenolic compounds. Grapes (Vitis) are among the fruits consumed in greatest quantities around the world, whether processed or in their natural form, and they also have one of the highest phenolic compound contents.2 Grape juice has also been reported to improve cognitive and motor function.3,4 Some of these effects are related to protection against oxidative stress,5 and the antioxidant activity of grape juices may be an indicator of the relative level of health benefit they offer. Phenolic compounds are secondary plant metabolites that play a key role in the sensory and nutritional quality of fruits, vegetables and other plants.6 These compounds and their antioxidant activity have long been associated with the beneficial effects of grapes and wines.7,39 Catechin and gallic acid act as free radical scavengers, and epicatechin has also demonstrated antibacterial activity and protective effects against membrane oxidation.6 Grape juices are rich in phenolic compounds and different studies have demonstrated that these substances possess biological activity related to health benefits for the consumers.8,9 The phenolic compounds in grape juices, mainly the flavonoids flavanols, flavonols and anthocyanins, are associated with improved health, along with other compounds which are not flavonoids, such as phenolic acids and the stilbene resveratrol.10-14 The flavonols are represented mainly by kaempferol, quercetin and myricetin and simple orthomethylated derivatives such asisorhamnetin, which have received considerable interest due to their antioxidant properties.15 Among the flavanols, (+)-catechin, (-)-epicatechin and procyanidins have gained attention due to their antioxidant, antimicrobial and bactericidal activity.16 The principal anthocyanins found in juices are malvidin, cyanidin, delphinidin, petunidin, peonidin and pelargonidin. The consumption of these anthocyanins is associated to biological activities, such as antioxidant capacity and prevention of cardiovascular diseases.17 Phenolic acids, such as gallic, caffeic and chlorogenic, have been studied for their antioxidant capacity and for acting as venous dilators.18 The aim of the present study is (1) to explore the potential antioxidant capacity, using the DPPH test, (2) to systematically evaluate the antioxidant capacity, total phenolic and flavonoid contents and to evaluate the changes in the activities of some antioxidant enzymes catalase, peroxidase and polyphenol oxidase in three locally growing grapes. This information will be useful for nutritionists and consumers.

MATERIALS AND METHODS

Grape samples

The study included three types of grape (Aasmi, Raziqi and Aswad) were obtained from three production areas/governorates (Sanaa, Amran and Saadah). Each sample was individually analyzed by triplicate.

Chemicals

The solvent used in the present work were purchased from Riedel-de-Haen (Germany). 1,1-Diphenyl-2-picrylhydrazyl (DPPH), 2,2’-azino-bis
Preparation of solvent extracts
Two g of grape peel or flesh were extracted by shaking at 150 rpm and 25°C for 24 h with 10 ml of 80% methanol and filtered through filter paper No. 1. The filtrate designated as methanol extract.

Determination of the total phenolic contents
Total phenolic content was measured according to available literature.\textsuperscript{16} Fifty µL of the methanol extract was mixed with 100 µL Folin-Ciocalteu reagent, 850 µL of distilled water and allowed to stand for 5 min at ambient temperature. A 500 µL of 20% sodium carbonate was added and allowed to react for 30 min. Absorbance was measured at 750 nm. Total phenols were quantified from a calibration curve obtained by measuring the absorbance of known concentrations of gallic acid (Figure 1). The results expressed as mg gallic acid equivalent/g tissues.

Determination of the total flavonoid contents
The total flavonoid content of the samples was determined using a modified colorimetric method described previously by\textsuperscript{17} and used catechin as a standard. Methanol extract or standard solution (250 µL) was mixed with distilled water (1.25 mL) and 5% NaNO\textsubscript{2} solution (75 µL). After standing for 6 min, the mixture was combined with 10% AlCl\textsubscript{3} solution (150 µL), 1 MNaOH (0.5 mL) and distilled water (275 µL) was added to the mixture 5 min later. The absorbance of the solutions at 510 nm was then measured. Total flavonoids were quantified from a calibration curve obtained by measuring the absorbance of known concentrations of gallic acid (Figure 2). The results expressed as mg catechin equivalent/g tissues.

Antioxidant assays

DPPH radical scavenging activity
Free radical scavenging activity of crude methanol extract was determined using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method. A methanol solution (100 µL) containing methanol extracts was added to 900 µL of freshly prepared DPPH methanol solution (0.1 mM). An equal amount of methanol was used as a control. After incubation for 30 min at room temperature in the dark, the absorbance was measured at 517 nm using a spectrophotometer. Activity of scavenging (%) was calculated using the following formula:

\[
\text{DPPH radical scavenging} = \left( \frac{\text{OD control} - \text{OD sample}}{\text{OD control}} \right) \times 100
\]

The results were plotted as the % of scavenging activity against concentration of the sample. The inhibition concentration (IC\textsubscript{50}) was defined as the amount of crude methanol extract required for 50% of free radical scavenging activity. The IC\textsubscript{50} value was calculated from the plots as the antioxidant concentration required for providing 50% free radical scavenging activity (Figure 2).

ABTS radical cation decolorization assay
ABTS (2,2’-azino-bis (3-ethylbenzo-thiazoline-6-sulfonic acid) also forms a relatively stable free radical, which decolorizes in its non-radical form. The spectrophotometric analysis of ABTS•+ scavenging activity was determined according to the method of Prieto et al.\textsuperscript{16} Sample solutions (50 µL) were combined in an Eppendorf tube with 1 mL of reagent solution (0.6 M sulfuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate), The tubes were capped and incubated in a thermal block at 95°C for 90 min. After the samples had cooled to room temperature, the absorbance of aqueous solutions of each was measured at 820 nm against a blank. The antioxidant activity was expressed as the absorbance of the sample (Figure 3).

Phosphomolybdenum complex assay
Spectrophotometric evaluation of antioxidant activity through the formation of a phosphomolybdenum complex was carried out according to Prieto et al.\textsuperscript{17} Sample solutions (100 µL) containing methanol extracts were added to 1 MNaOH (0.5 mL) and distilled water (275 µL) was added to the mixture 5 min later. The absorbance of the solutions at 750 nm was then measured. Total flavonoids were quantified from a calibration curve obtained by measuring the absorbance of known concentrations of gallic acid (Figure 1). The results expressed as mg catechin equivalent/g tissues.
supernatant was designed as crude extract and stored at −20°C for further analysis.

**Peroxidase assay**

Peroxidase activity was carried out according to Yuan and Jiang. The reaction mixture contained in one ml: 8 mM H2O2, 40 mM guaiacol, 50 mM sodium acetate buffer, pH 5.5 and least amount of crude extract. The change in absorbance at 470 nm due to guaiacol oxidation was followed for 1 min using a spectrophotometer. One unit of peroxidase activity was defined as the amount of enzyme which increased the O.D. 1.0 per min under standard assay conditions.

**Catalase assay**

Catalase activity was determined according to Bergmeyer. Two and half ml of substrate solution was made up of 25 mM H2O2 in a 75 mM sodium phosphate buffer pH 7.0 and crude extract. The decrease in absorbance at 240 nm and 25°C was recorded for 1 min using a spectrophotometer. One unit of enzyme activity was defined as the amount of the enzyme that causes a change of 0.1 in absorbance per min under standard assay conditions.

**Polyphenoloxidase assay**

Polyphenoloxidase activity was assayed with catechol as a substrate according to the spectrophotometric procedure of Jiang et al. The enzyme solution (100 μL) was rapidly added to 900 μL of 40 mM catechol solution prepared in 0.01 M sodium phosphate buffer, pH 6.8. The increase in absorbance at 400 nm and 25°C was recorded for 3 min using a spectrophotometer. One unit of enzyme activity was defined as the amount of the enzyme that causes a change of 0.1 in absorbance per min.

**Statistical analysis**

The statistical analyses were performed by a one-way ANOVA and the Student's t-test. The results were expressed as means ± S.E. to show variations in the various experimental. Difference are considered significant when \( P < 0.05 \).

**RESULTS**

**Estimation of the total content of phenols**

The results for the total content of phenolics of the grapes from 3 types (Aasmi, Aswad, Raziqi) are giving in Table 1. A great variation in terms of total phenolic content was observed among the grape cultivars (2.5-7.4 mg GAE/g peel; 0.33-1.32 mg GAE/g flesh). The highest amount of total phenolics in peel was observed for Aasmi (7.4 mg GAE/g peel), followed by Aswad (5.3 mg GAE/g peel), while lowest was in Raziqi (2.5 mg GAE/g peel). For flesh, the highest amount of total phenolics was also observed for Aasmi (1.32 mg GAE/g flesh), followed by Raziqi (0.72 mg GAE/g flesh), while lowest was in Aswad (0.33 mg GAE/g flesh). According to the literature, the total phenolic compounds concentrations in commercial grape juices in Brazil varies from 270 to 3433 mg/L, with mean values of between 1430 and 1915 mg/ L, these differences being mainly due to the production techniques and to the particular
characteristics of the region. In commercial Spanish juices reported values of 705 to 1177 mg/L, while the values for juices produced from the varieties Noble (V. rotundifolia) and Concord (V. labruscana) varied from 1280 to 2880 mg/L, respectively.

Estimation of the total content of flavonoids

The total content of flavonoids in grape from 3 types (Aasmi, Aswad, Raziqi) ranged from 0.97 to 1.94 mg CE/g peel and 0.08 to 0.46 mg CE/g flesh) (Table 2). The highest amount of total flavonoids in peel was observed for Aasmi (1.94 mg CE/g peel), followed by Aswad (1.38 mg CE/g peel), while lowest was in Raziqi (0.97 mg CE/g peel). For flesh, the highest amount of total flavonoids was also observed for Aasmi (0.46 mg CE/g flesh), followed by Raziqi (0.31 mg CE/g flesh), while lowest was in Aswad (0.08 mg GAE/g flesh). The concentration of catechin in the red grape juices was within the range of previous reports in this type of juice (0.38–86.4 mg/L). Epicatechin concentrations in the grape juices were also within the range of previous findings (from 0.92 to 22.13 mg/L) in grape juices, whereas the concentrations in the white and red wines were lower than previous reports (from 0.68 to 150 mg/L). Catechin acts against free radicals and chelating metals, and epicatechin has demonstrated vasodilator activity in human beings.

DISCUSSION

The phenolics content of grape from different cultivars showed concentration dependent scavenging of DPPH radical, which may be attributed to its hydrogen donating ability (Figure 1). For all grape cultivars tested, a linear increase of antioxidant activity was detected with increasing the scavenging IC<sub>50</sub> value of the test sample that decrease 50% initial radical.

Table 3: Antioxidant effect of phenolic concentrations of different grape cultivars on reduction of DPPH radical scavenging

<table>
<thead>
<tr>
<th>Grape cultivar</th>
<th>Tissue</th>
<th>IC&lt;sub&gt;50&lt;/sub&gt; (µg phenolic content)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aasmi</td>
<td>Peel</td>
<td>10.2</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>5.8</td>
</tr>
<tr>
<td>Raziqi</td>
<td>Peel</td>
<td>5.7</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>3.9</td>
</tr>
<tr>
<td>Aswad</td>
<td>Peel</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>1.7</td>
</tr>
</tbody>
</table>

IC<sub>50</sub> (µg phenolic content) is the inhibition concentration of the test sample that decrease 50% initial radical.

Table 4: Antioxidant effect of phenolic concentration of different apple cultivars on reduction of ABTS radical scavenging

<table>
<thead>
<tr>
<th>Apple cultivar</th>
<th>Tissue</th>
<th>IC&lt;sub&gt;50&lt;/sub&gt; (µg phenolic content)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aasmi</td>
<td>Peel</td>
<td>3.2</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>1.8</td>
</tr>
<tr>
<td>Raziqi</td>
<td>Peel</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>1.7</td>
</tr>
<tr>
<td>Aswad</td>
<td>Peel</td>
<td>1.37</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>1.7</td>
</tr>
</tbody>
</table>

IC<sub>50</sub> (µg phenolic content) is the inhibition concentration of the test sample that decrease 50% initial radical.

Table 5: Antioxidant effect of phenolic concentration of different apple cultivars on formation of phosphomolybdenum complex

<table>
<thead>
<tr>
<th>grape cultivar</th>
<th>Tissue</th>
<th>EC&lt;sub&gt;50&lt;/sub&gt; (µg phenolic compounds)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aasmi</td>
<td>Peel</td>
<td>9.8</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>1.37</td>
</tr>
<tr>
<td>Raziqi</td>
<td>Peel</td>
<td>2.9</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>0.79</td>
</tr>
<tr>
<td>Aswad</td>
<td>Peel</td>
<td>5.9</td>
</tr>
<tr>
<td></td>
<td>flesh</td>
<td>0.76</td>
</tr>
</tbody>
</table>

EC<sub>50</sub> (µg phenolic content) is the efficient concentration of the test sample that increases O.D 0.5.

Table 6: The antioxidant enzyme activities of different grape cultivars

<table>
<thead>
<tr>
<th>grape cultivar</th>
<th>Tissue</th>
<th>Units of polyphenol oxidase/g tissues</th>
<th>Units of peroxidase/g tissues</th>
<th>Units of catalase/g tissues</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aasmi</td>
<td>Peel</td>
<td>148 ± 0.05</td>
<td>44 ± 0.05</td>
<td>18 ± 0.03</td>
</tr>
<tr>
<td></td>
<td>Flesh</td>
<td>65 ± 0.03</td>
<td>-</td>
<td>9 ± 0.007</td>
</tr>
<tr>
<td>Raziqi</td>
<td>Peel</td>
<td>87 ± 0.07</td>
<td>38 ± 0.012</td>
<td>14 ± 0.003</td>
</tr>
<tr>
<td></td>
<td>Flesh</td>
<td>20 ± 0.02</td>
<td>-</td>
<td>7 ± 0.06</td>
</tr>
<tr>
<td>Aswad</td>
<td>Peel</td>
<td>32 ± 0.06</td>
<td>34 ± 0.08</td>
<td>9 ± 0.017</td>
</tr>
<tr>
<td></td>
<td>Flesh</td>
<td>15 ± 0.008</td>
<td>-</td>
<td>4 ± 0.006</td>
</tr>
</tbody>
</table>
phenolic concentrations. In grape cultivar peels, the correlation coefficient (R²) between phenolic concentrations and DPPH scavenging activity was found to be 0.99 (Raziqi), 0.97 (Aswad), 0.90 (Aasmi). In flesh, the correlation coefficient (R²) between phenolic concentrations and DPPH scavenging activity was found to be 0.99 (Aasmi), 0.97 (Raziqi), 0.96 (Aswad). These high values of correlation coefficient (R²) indicating the strong correlation. The DPPH assay IC₅₀ value (the inhibition concentration as µg phenolic content of the test sample that decrease 50% initial radical) was ranged from 5.7 to 10.2 µg phenolic concentrations/ml grape cultivar peels (Table 3). The highest IC₅₀ was observed for grape peels of Raziqi (5.7 µg phenolic concentration), followed Aswad (6 µg phenolic concentration), while the lowest was in Aasmi(10.2 µg phenolic concentration). For grape fleshes, the highest IC₅₀ was also observed for Aasmi(1.7 µg phenolic concentration), followed Raziqi (3.9 µg phenolic concentration), while the lowest was in Aasmi (5.8 µg phenolic concentration).

The phenolics content of grape from different cultivars showed concentration dependent scavenging of ABTS radical, which may be attributed to its hydrogen donating ability (Figure 2). For all grape cultivar tested, a linear increase of antioxidant activity was detected with increasing the phenolic concentrations. In grape cultivar peels, the correlation coefficient (R²) between phenolic concentrations and ABTS scavenging activity was found to be 0.97 (Aswad), 0.94 (Raziqi), 0.93 (Aasmi). In flesh, the correlation coefficient (R²) between phenolic concentrations and ABTS scavenging activity was found to be 0.99 (Raziqi), 0.97 (Aasmi), 0.96 (Aswad). These high values of correlation coefficient (R²) indicating the strong correlation. The ABTS assay IC₅₀ value (is the efficient concentration that decreases 50% of initial absorbance) was ranged from 1.7 to 6 µg phenolic concentration/ml grape cultivar peels (Table 4). The highest IC₅₀ was observed for grape peels of Aswad (6 µg phenolic concentration), followed Aasmi (3.2 µg phenolic concentration), while the lowest was in Raziqi (1.7 µg phenolic concentration). For grape fleshes, the highest IC₅₀ was also observed for Aasmi (1.8 µg phenolic concentration), followed Aswad (1.7 µg phenolic concentration), while the lowest was in Raziqi (1.37 µg phenolic concentration).

Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolydnum complex showed an increase of activity with increasing the phenolic concentrations of grape cultivars (Figure 2). In grape cultivar peels, the correlation coefficient (R²) between phenolic concentrations and the formation of a phosphomolydnum complex was found to be 0.99 (Aasmi), 0.98 (Aswad), 0.98 (Raziqi). In flesh, the correlation coefficient (R²) between phenolic concentrations and the formation of a phosphomolydnum complex was found to be 0.99 (Aasmi), 0.98 (Raziqi), 0.90 (Aswad). These high values of correlation coefficient (R²) indicating the strong correlation. The formation of a phosphomolydnum complex EC₅₀ value (is the efficient concentration as µg phenolic content of the test sample that increases O.D. 0.5) was ranged from 0.76 to 9.8 µg phenolic concentration/ml grape cultivar peels (Table 5). The highest EC₅₀ was observed for grape peels of Raziqi (2.9 µg phenolic concentration), followed Aswad (5.9 µg phenolic concentration), while the lowest was in Aasmi Raziqi (9.8 µg phenolic concentration). For grape fleshes, the highest EC₅₀ was observed for Aswad (0.76 µg phenolic concentration), followed Raziqi (0.79 µg phenolic concentration), while the lowest was in Aasmi (1.37 µg phenolic concentration).

Table 6 shows the activities antioxidant enzymes, polyphenoloxidase, peroxidase and catalase in the crude extracts of the peel and flesh of the grape cultivars (Aasmi, Raziqi and Aswad). The highest level of polyphenoloxidase activity was recorded in the peel of Aasmi, Raziqi and Aswad with 148, 84 and 32 units/g tissues, respectively. Moderate level of peroxidase activity was observed for all grape cultivars tested (34 to 44 units/g tissues). All apple cultivars tested exhibited lowest level of catalase activity (9 to 18 units/g tissues).

CONCLUSION

The results indicated that the imported grapes retained the most antioxidant activities. The present manuscript precisely explores the beneficial effect of yemeni grapes which are very rich source of antioxidants. These grapes contains high amount of free radicals scavengers which are intact and thus are helpful in the delaying aging process, cancer, and other related pathophysiological diseases which are so common in today's stressed lifestyle. This manuscript reveals the enzymes present in yemeni grapes that are responsible to combat and prevent the diseases. This manuscript in current light of research promotes the cultivation and consumption of yemeni grapes for the general good to consumers, nutritionist thus boosting the economy of grapes cultivators. Finally, this study will be supply information on the antioxidant activity of grapes that benefits grapes producers, nutritionists and consumers.

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CONFLICT OF INTEREST

The authors report no conflict of interest.

ABBREVIATION USED

DPPH: 2,2-diphenyl-1-picrylhydrazyl; ABTS: 3-ethylbenzo-thiazoline-6-sulfonic acid; FAO: Food and Agriculture Organization; IC₅₀: Inhibition concentration; R²: Correlation coefficient.

REFERENCES

SUMMARY

- The phenolic content of peel and flesh from different cultivars of grapes showed concentration dependent scavenging of DPPH and ABTS radicals.
- The phenolic content of grapes revealed the formation of phosphate/molybdenum complex through which extract perform its scavenging activity.
- High correlation coefficient (R²) was observed between yemini grapes and its antioxidant activity.
- The phenolic extract revealed the high amount of polyphenoloxidase, peroxidase and catalase activity.
- The extract revealed the presence of total flavonoids calibrated with catechin acid.

ABOUT AUTHORS

Ruqaiyah Khan: Obtained her PhD degree in the field of Pharmacology focused on liver cancer and cardiac remodeling. She have excellent skill in paper and project writing with significant novel ideas. Moreover, she has many international publications on her name and has command over experimental pharmacology concerned with liver cancer. She is currently appointed as associate professor at Siddhartha Institute of Pharmacy and an active member of Institutional Animal Ethical committee.

Firoz Anwar: He has experience of more than 20 years in academics and research. Currently he is working as professor in King Saud University, Jeddah Saudi Arabia. He was former Assistant Director of All India Council for Technical Education, India. He has excellent command over Cancer biology, Microbiology, Toxicology and Pharmacology field. He is a potential reviewer of many renowned journals like The Lancet, etc. He is having more than 60 international publications and some patents to his name. He is expert in making protocols with significant novel ideas in the field of pharmacology, pharmacognosy. He has successfully guided doctorate students to attain their PhD degrees.