Clinical Attestation and Redox Mechanism of Ascorbic Acid in the Treatment of Cancer

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ABSTRACT

Ascorbic acid (Vitamin C) is an essential micro-nutrient, an outstanding antioxidant and an essential co-factor in different mammalian enzymatic processes. There is considerable clinical attestation that, high dose of intravenous ascorbic acid can improve cancer patients with or without extant therapeutic involvements. Ascorbic acid in high intravenous doses serves as pro-oxidant and promotes the generation of hydrogen peroxide (H_2O_2) and other reactive oxygen species (ROS) with oxidative stress-induced toxicity selectively to cancer cells. This effect also hampers the bioenergetics and angiogenesis of malignant cells, resulting in cancer cell death. Large doses of ascorbic acid are safe and well-tolerated. On account of its antioxidant effect, ascorbic acid supplementation may be applied as an adjuvant with regular cancer therapy to minimize complications. Nevertheless, there is a necessity for further mechanistic studies and randomized controlled clinical trials to evaluate the benefit of ascorbic acid in the treatment of cancer.

Key words: Ascorbic acid, Reactive oxygen species (ROS), Intravenous, Angiogenesis, Clinical.

INTRODUCTION

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L-ascorbic acid (and its salts - ascorbate), commonly known as vitamin C, is a water-soluble vitamin micronutrient. Unlike most mammals and other animals, humans cannot synthesize this vitamin hence, should acquire it from the food. It is a natural product (phytochemical) found in higher plants – several fruits and vegetables and can also be synthesized. It was used for prevention and treatment of scurvy. Ascorbic acid (ascorbate) is an essential co-factor in several enzymatic biochemical reactions viz. biosynthesis of collagen, carnitine, and neuropeptides (including certain neurotransmitters) and in regulation of gene expression and immune system functions. It is also an excellent natural antioxidant.¹

The anticancer effect of ascorbic acid has been appearing from more than the last six decades. In 1950s, there was a hypothesis that ascorbic acid exercises anticancer activity by promoting collagen synthesis.^{2,3} In 1972, the next hypothesis suggested that, the anticancer activity of ascorbic acid was owing to inhibition of hyaluronidase.⁴ The cytotoxic property of ascorbic acid was first reported in 1969, when sodium ascorbate was demonstrated to effect death of Ehrlich ascites carcinoma cells *in vitro*.⁵ The applications of intravenous ascorbic acid in cancer treatment had been investigated since the 1970s.^{6,7} Since then, ascorbic acid was receiving attention in management of human cancers. Numerous experimental pre-clinical and clinical studies put forward the favorable effects of ascorbic acid in prevention and treatment of different human cancers exhibiting oxidation-reduction (redox) mechanisms. The present review collates the noteworthy clinical attestations of cancer treatment with ascorbic acid and endeavors to critically explicate the underlying redox mechanisms.

PHYSIOLOGICAL REDOX ROLES

The prescribed daily dietary requirement of vitamin C is 75-90 mg. Ascorbic acid is a potent antioxidant or reducing agent, i.e., it donates electrons to recipient molecules – as per electronic redox concept. Due to this redox effect, there are two chief physiological roles of ascorbic acid: as an antioxidant and as an enzyme co-factor.^{1,8}

Ascorbic acid is the non-enzymatic antioxidant systematically prevalent in blood and tissues. The normal blood plasma concentration of ascorbic acid is 50-100 μ M in healthy adults. Owing to antioxidant property, it is able to rapidly undergo gradual loss of one or two electrons i.e., successive oxidations, producing the relatively stable ascorbyl/ascorbate free radical (semidehydroascorbic acid) or unstable dehydroascorbic acid (DHA), respectively (Figure 1). Ascorbate radical can be reduced to ascorbic acid or may further be oxidized to DHA. In cells, DHA is readily reduced by glutathione

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Figure 1: Oxidation of ascorbic acid.

Table 1: The major enzymes requiring ascorbic acid as co-factor in mammals.

SI. No.	Enzyme types	Enzymes	Functions
1	Monooxygenase	Dopamine β-monooxygenase	Noradrenaline biosynthesis
2		Peptidyl-glycine α amidating monooxygenase	Amidation of peptide hormones
3	Dioxygenase	3 Prolyl 3-hydroxylase isoenzymes	Collagen biosynthesis
4		3 Lysyl hydroxylase isoenzymes	
5		4 Hypoxia-inducible factor (HIF) isoenzymes	HIF hydroxylation
6		γ-Butyrobetaine hyroxylase	Carnitine biosynthesis
7		4-Hydroxyphenylpyruvate dioxygenase	Tyrosine metabolism
8		10-11-translocation family of dioxygenases	Demethylation of DNA

or thioredoxin to ascorbic acid. In this manner, ascorbic acid guards vital cellular bio-macromolecules like proteins, lipids and nucleic acids (DNA and RNA), from oxidative decomposition by free radicals/reactive oxygen species (ROS) that are produced on normal metabolic conditions and overproduced in disease conditions and invoked by toxins, xenobiotics (like drugs) and environmental contaminants. Ascorbic acid also takes part in redox salvage of other putative antioxidants, like regeneration of tocopherol (Vitamin E) from its oxidized state.⁹

The function of ascorbic acid as co-factor is also associated with its redox role. Ascorbate can chelate and reduce transition metal ions especially Fe3+ and Cu2+. Owing to this capability, ascorbate serves as an essential co-factor for several iron and copper-bearing enzymes. Ascorbate acts as co-factor for about 150 (one hundred fifty) human enzymes. By keeping enzyme-bound metals in their reduced state, ascorbic acid helps mixed-function oxidases for the synthesis of several prime biomolecules.1 These pertinent enzymes are either monooxygenases or dioxygenases (Table 1). Symptoms of Vitamin C deficiency, like slow wound healing and lethargy, cause from the disablement of these ascorbic acid-dependent enzymatic reactions resulting in the inadequate synthesis of catecholamines, carnitine, collagen etc. Furthermore, different dioxygenases are associated in the regulation of gene expression and the preservation of genome require ascorbic acid as a co-factor. For instance, ascorbate is entailed in the regulation of stability of hypoxia inducible factors (HIF) which mediate angiogenesis.¹⁰

CLINICAL ATTESTATIONS IN CANCER TREATMENT

Experiments performed in the 1970s revealed that, high doses (quite higher than normal dietary requirement) of intravenous ascorbic acid (10 g/day infused intravenously for 10 days) were helpful in extending the life span and improving the quality of life of terminal cancer patients.67 Concomitant clinical studies with same high dose (10 g/day) ascorbic acid given orally to advanced stage cancer victims however, demonstrated no significant increase in survival time.^{11,12} Therefore, is evident that, the intravenous route of ascorbic acid administration is crucial to attain this beneficial action. Compared to ascorbic acid administered orally, intravenous ascorbic acid can produce 30-70 times more plasma ascorbic acid concentrations.13,14 Outcome from recently conducted controlled clinical trials demonstrated that, intravenous ascorbic acid was safe and well tolerated by the cancer patients. There is no testimonial that, high dose of ascorbic acid (up to 10 g/day) impose any adverse or toxic reactions.^{15,16} Notwithstanding, while generally regarded as a nutraceutical or dietary supplement, neither the U.S. Food & Drug Administration (US-FDA) nor European Medicines Agency has approved the usage of intravenous high dose ascorbic acid in the treatment of cancer.17

Further succeeding clinical investigations demonstrated that, combination of high dose ascorbic acid with conventional anticancer therapies retards cancer growth in human models of pancreatic,^{18,19} hepatic,²⁰ breast,²¹ metastatic pancreatic cancer,²² sarcoma and malignant mesothelioma.²³ The momentous prototypical clinical trials using intravenous ascorbic acid in treatment of cancer or obviation of anticancer treatment-invoked complications are summarized at Table 2. There is no proof that cancer advancement occurs with intravenous ascorbic acid supplementation. The recent study of 2018 is the evidence of randomized controlled phase II trial which seems to be the most motivating beneficial effect i.e., remission found in elderly acute myeloid leukemia patients in combination with chemotherapy (decitabine etc.), exhibiting a synergistic outcome.²⁴ Few clinical studies using oral ascorbic acid administration till date yielded no response as stated in the foregoing paragraph, regardless of interventions.

Clinical trials of complementary high dose intravenous ascorbic acid infusion in cancer patients have improved the quality of life (QOL), as well as developments in physical and mental states and melioration of conventional cancer therapy-induced adverse events namely nausea, vomiting, fatigue, pain; hair, weight and appetite loss etc.^{10,21,22,25} Moreover, ascorbic acid has also reported to abrogate the harmful effects of radiation therapy²⁶ and chemotherapeutic agents like bleomycin,¹⁷ tamoxifen²⁷ etc when given by other routes (like oral) in low/pharmacological doses, causing decreased morbidity.²⁸

Literature haunt unveils that, current attestations of the efficacy of intravenous ascorbic acid in cancer patients is restricted to observational studies, uncontrolled interventions, phase I/II studies and case reports. Interventions and patient population are divergent. The clinical efficacy should be determined exhaustively in specific cancer types or populations. Hence, there is a necessity for large, longer-duration, randomized controlled phase II clinical trials with proper controls that assess the efficacy of intravenous ascorbic acid in progression of specific cancers and survival thereof.¹⁶

REDOX MECHANISM OF ANTICANCER EFFECT

The mechanism of anticancer effect of intravenous ascorbic acid is still under critical exploration. Even though physiologically ascorbic acid serves as antioxidant, its therapeutic effectiveness at pharmacological or

Table 2: Summary of important clinical studies on intravenous ascorbic acid in the treatment of cancer.

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SI. No.	Year reported	Study type	Dose (g)	Regimen	Population type, number of patients	Concomitant chemotherapy/ radiation/ surgery	Efficacy	Reference no.
1	1976	Observational	10	Daily for 10 days	Advanced cancer, 100	None	OS prolonged, 4 times	6
2	1978	Observational	10	Daily for 10 days	Advanced cancer, 100	None	OS prolonged 4.6 times	7
3	1982	Observational	10-20	Daily	Advanced cancer, 111	None	OS prolonged 5.7 times, improved QOL	29
4	1991	Observational	10-45	Several	Advanced cancer, 294	Yes	OS prolonged 1.9 times	30
5	2005	Phase I	10-50	Daily for 8 weeks	Advanced cancer, 294	None	Stable	31
6	2007	Observational	10	Twice with 3 days interval	Stage IV cancer, 39	None	Improved QOL	28
7	2008	Phase I	0.4-1.5/ kg	Thrice in a week for 2 weeks	Solid and leukemic, 24	None	Stable, Improved QOL	32
8	2011	Observational	7.5	Weekly for 4 weeks	Stage II breast cancer, 53	Yes	Improved QOL 2 times	21
9	2012	Observational	50	Thrice in a week, median 9 treatments	Prostate and other cancers, 45	None	Decreased PSA and CRP	33
10		Observational	50	Twice in a week for 4 weeks	Advanced cancer, 60	None	Improved QOL	34
11		Phase I	50-100	Thrice in a week for 8 weeks	Stage IV pancreatic cancer, 9	Gemcitabine and Erlotinib	Tumor mass reduced, OS prolonged	22
12	2013	Phase I/IIA	50-125	Twice in a week for 4 weeks	Stage IV pancreatic cancer, 9	Gemcitabine	Improved QOL, OS prolonged	35
13		Phase I	50-110/ m ²	Four times in a week for 4 weeks	Solid tumors, 17	Ecosapentanoic acid	Stable	20
14	2014	Phase I/IIA	75-100	Twice/thrice in a week for 1 year	Stage III-IV ovarian cancer, 27	Paclitaxel and carboplatin	Prolongation of relapse	36
15	2015	Phase IIA: uncontrolled	86-124 (1.5 g/kg)	Thrice in a week for 11- 580 days	Advanced cancer, 14	Yes	Stable, partly long lasting disease	37
16	2017	Phase II: uncontrolled	5-60	Weekly for 12 weeks	Metastatic castration-resistant prostate cancer, 23	None	Marginal improvement of QOL	38
17	2018	Phase IIB: randomized controlled	50–80 mg/ kg	10 Days per month, for 10 Months	Acute myeloid leukemia, 39	Decitabine, cytarabine and aclarubicin	OS prolonged 1.6 times, remission rate higher	24

Notes: OS: Overall survival; QOL: Quality of life; PSA: Prostate specific antigen; CRP: C-reactive protein.



Figure 2: Fenton reaction of ascorbic acid.

higher doses in most cancer cases seems to be connected to pro-oxidant effects eventually causing cancer cell death.^{14,39}

The recent mechanistic studies are based on reactive oxygen species (ROS) induced cellular events, because the generation of hydrogen peroxide (H_2O_2) is cardinal event in ascorbate-induced anticancer effect; and H_2O_2 is a ROS that can generate ROS down-stream through transmetal catalyzed Fenton reaction.^{14,39,40} This reaction is the oxidation of organic substrates such as ascorbic acid by iron (III) to produce H_2O_2 (Figure 2).

1. Ascorbic acid (AscH₂) reduces ferric ions (Fe³⁺) to ferrous ions (Fe²⁺).

2. Ferrous ion reacts with oxygen to produce superoxide radical.

3. Dismutation of superoxide generates hydrogen peroxide.

Ascorbic acid selectively ruins cancer cells through the intracellular production of toxic H2O2 generated upon its oxidation through aforesaid scheme. High doses of ascorbic acid after intravenous administration serves as a peroxide delivery system (pro-drug) for the generation of stable ascorbate radical and H2O2, with concurrent selective oxidative damage of cancer cells.^{40,41} Cancer cells, as compared to normal ones, show impaired endogenous enzymatic antioxidant mechanism consisting of superoxide dismutase, catalase, glutathione S-transferase, glutathione peroxidase etc. The reduced activity of antioxidant enzymes hinders further degradation of H₂O₂ thus causing intracellular accumulation of H₂O₂ (ROS) which in turn, generate more ROS downstream as per the chain reactions leading to intracellular redox imbalance and oxidative impact resulting in oxidative damage to cellular vital macromolecules and organelle. Normal cells having competent endogenous enzymatic defense system, detoxifies H₂O₂ regularly - therefore, no or less cellular accumulation and resultant cytotoxicity.40-42

In human body, ascorbic acid is oxidized to dehydroascorbic acid (DHA) which is then reduced further to ascorbic acid within the cells (see above). The reduction of DHA to ascorbic acid scavenges reduced glutathione (GSH). It also acts as reducing agent (antioxidant) when it detoxifies hydrogen peroxide (ROS) by means of the enzyme glutathione peroxidise. GSH thus functions as the cellular non-enzymatic antioxidant defence as it abrogates oxidative stress by reducing ROS. Hence, depletion of GSH content invokes weakening of non-enzymatic redox mechanism and elicits oxidative impact leading to death of cancer cells.^{43,44}

Recent studies suggest a salient relationship between ascorbic acid and GSH entailing glucose metabolism including glycolysis, tricarboxylic acid cycle (TCA) or citric acid cycle and pentose phosphate pathway. Ascorbate-induced oxidative stress with depletion of GSH, results in inhibition of a glycolytic enzyme viz. glyceraldehyde-3-phosphate-dehydrogenase (GAPDH).^{44,45} Therefore, the upstream GAPDH metabolites i.e., metabolites associated with upstream glycolysis, partial TCA cycle (e.g., citrate and cis-aconitate) and pentose phosphate pathway accumulate whereas downstream metabolites of GAPDH are degraded except citrate and cis-aconitate. High dose of ascorbic acid reduces the synthesis of ATP, as high dose ascorbic acid interferes with glycolysis and TCA cycle energy metabolism, consequently restricting ATP production. Ascorbic acid-mediated oxidative insult, in turn, induces the depletion of NADH, which also obstructs glycolytic pathway. The decreased ATP level leads to intracellular energy crisis which results in cancer cell death.^{46,47}

In accordance with this mechanism, ascorbic acid acting as a pro-oxidant, increases the intracellular reactive oxygen species (ROS), which leads to genotoxicity i.e., DNA damage, with consequent activation of poly ADP-ribose polymerase (PARP), an enzyme required for DNA repair. PARP activation in turn quenches NAD⁺ with NAD⁺ depletion and resultant ADP degradation, leading to energetic dearth and death of malignant cells.^{47,48}

Due to mitochondrial dysfunction and lack of adequate oxygen supply i.e., hypoxia in the tumor microenvironment owing to deficient blood circulation, cancer cells live mainly on glycolysis for ATP production, hence their ATP synthesis is not efficient as compared with normal cells which generally utilize oxidative phosphorylation.^{49,50} Therefore, the cancer cells become more vulnerable to ascorbate-induced oxidative stress-mediated energetic paucity than normal cells.⁵¹ Nevertheless, the selective anticancer action of ascorbic acid against cancer cells and its mechanisms require further investigations.

Ascorbic acid and DHA-induced perturbations in the cellular redox system decreases the available nitric oxide (NO) in endothelial tissues. NO is regarded as a main impetus for new blood vessel formation i.e., angiogenesis. Angiogenesis is the pertinent event for most of the solid tumors to get rid of hypoxia. NO pathways are the key elicitors of tumor angiogenesis. High concentrations of ascorbates impedes the generation of NO therefore; high concentrations of ascorbic acid have been demonstrated to prevent angiogenesis. Intracellular downregulation of HIF (which is generally upregulated in solid tumors) by ascorbic acid can also induce angiostasis. Ascorbic acid is thus found to limit tumor angiogenesis, resulting in prevention of carcinogenesis in experimental animals.^{17,52,53}

The role of reactive oxygen species (ROS) in pathogenesis of cancer has duly been established.⁵⁴ Physiological levels of ascorbic acid effectively detoxify ROS that are produced during normal metabolism and overproduced under different types of stress and disease conditions owing to antioxidant effect.⁵⁵ Therefore, ascorbic acid protects tissue damage and death induced by pro-oxidant stressors viz. xenobiotics and pollutants.⁹ It is apparent that, alleviation in toxicity of extant chemotherapy and/ or radiotherapy on normal tissue upon complementary co-treatment with ascorbic acid is associated with abrogation of concomitant oxidative injury to non-target normal cells thereby improving the life quality of patients receiving chemo/radiotherapy, demonstrating its benefit to be applied as adjuvant or palliative treatment.^{56,57}

CONCLUSION

Ascorbic acid, a phytochemical (natural product), is an essential vitamin micro-nutrient, a potent antioxidant and essential co-factor in several major mammalian enzymes. There is significant clinical proof that, high dose intravenous ascorbic acid can improve the condition of cancer patients with or without typical therapeutic interventions. The effect is observed particularly with intravenous than oral application. High dose intravenous ascorbic acid is found safe and well endured in cancer patients. Nonetheless, the clinical efficacy requires to be assessed congruently in specific types of cancer. Hence, there is need for definitively designed randomized controlled clinical trials to evaluate the efficacy of intravenous ascorbic acid in treatment of neoplastic diseases. Ascorbic acid supplementation may also be employed as adjuvant with conventional cancer therapies to circumvent corresponding complications. The clinical data attest to conclude that, ascorbic acid possesses the prospect to be an effective, innocuous, easily available cancer therapy in due course. Mechanistically, ascorbic acid at large intravenous doses, acts as a pro-oxidant and promotes intracellular hydrogen peroxide (H₂O₂) and other ROS generation leading oxidative stress-mediated selective aftermath to malignant cells. This effect also interferes with bioenergetics

and angiogenesis of cancer cells, causing cancer cell death. The molecular mechanisms of its selective actions against cancer cells warrant advanced studies. Further clinical as well as mechanistic studies are the need of the hour to reap the absolute benefits of this vitamin in cancer therapy.

CONFLICT OF INTEREST

The author declares no conflict of interest.

ABBREVIATIONS

ROS: Reactive oxygen species; **ATP:** Adenosine triphosphate; **ADP:** Adenosine diphosphate; **DHA:** Dehydroascorbic acid; **NADH:** Nicotinamide adenine dinucleotide (reduced); **PARP:** Poly ADP-ribose polymerase; **DNA:** Deoxyribonucleic acid; **RNA:** Ribonucleic acid; **QOL:** Quality of life; H_2O_2 : Hydrogen peroxide; **GAPDH:** Glyceraldehyde-3-phosphate-dehydrogenase; **GSH:** Reduced glutathione.

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